Distribution and Abundance of Cephalopod Paralarvae in the Bay of Bengal

Natinee Sukramongkol, Ritthirong Prommas and Sayan Promjinda

Southeast Asian Fisheries Development Center, Training Department P.O.Box 97, Phrasamutchedi, Samutprakarn 10290, THAILAND

Abstract

The 34 survey stations were conducted to collect the cephalopod paralarvae in the Bay of Bengal aboard M.V. SEAFDEC during 6 November to 7 December 2007 using a pair of bongo nets. A total of 278 paralarvae of 13 families and 19 genera were collected from 29 of 34 stations. The most numerous families were the Ommastrephidae (41% of total catch) followed by families Enoploteuthidae (14%) and Onychoteuthidae (6%). The majority of individuals were *Nototodarus hawaiiensis* (49 individuals) and *Abraliopsis* sp. (24 individuals). The paralarvae catch during the survey are all oceanic species (suborder Oegopsida) except 2 families from suborder Incirrata (order Octopoda). The compositions of the cephalopod paralarvae from the results show that the abundant paralarval squid is the commercial and/or minor-commercial target species to fisheries (Ommastrephidae, Enoploteuthidae, Thysanoteuthidae, Octopodidae).

Key words: cephalopod paralarvae, Bay of Bengal, distribution, abundance

Introduction

The composition and distribution of the cephalopod early life stages or 'paralarvae' (Young and Harman, 1988) has been investigated in the Pacific coast of Japan (Okutani, 1968 and 1969; Yamamoto and Okutani, 1975; Saito and Kubodera, 1993), the Gulf of Guinea (Arkhipkin et al., 1988), the Northwest Pacific (Okutani, 1966; Kubodera and Okutani, 1981; Kubodera and Jefferts, 1984a; Kubodera and Jefferts, 1984b; Kubodera, 1991), the Arabian Sea (Nesis, 1974; Piatkowski and Welsch, 1991; Piatkowski et al., 1993), and California current (Okutani and Mcgowan, 1969). Although, the Bay of Bengal is one of the largest marine ecosystem, the investigation of paralarvae distribution in the area is still sparse. The knowledge of the early life stages cephalopod distribution and abundance patterns is useful to determining the spawning area and period (Bower et al., 1999). It can also help in understanding cephalopod population dynamics and in developing stock-recruitment models for commercial important species (Vecchione, 1987). The joint research survey by the fishery sector of BIMSTEC to observe and collect scientific data concerned with fishery and oceanographic aspects in 2007 provided a valuable opportunity to analyze the data from the bongo net collection. The present study is objective to provide the information on the composition and distribution of young stages of cephalopod in an extensive region of the Bay of Bengal.

Materials and methods

The sampling of cephalopod paralarvae at 34 survey stations were conducted in three areas off Bangladesh waters (area A; between latitude 16[°]N-19[°]N, longitude 88[°]E-91[°]E),

India and Sri Lanka waters (area B; between latitude 09°N-14°N, longitude 82°E-85°E), and Myanmar waters (area C; latitude 10°N-12°N, longitude 95°E-97°E) (Fig. 1).

The standard sampling procedure was conducted aboard M.V. SEAFDEC during 6 November to 7 December 2007. All oblique tows made during the day of about 30 minute duration were taken at each station with a pair of bongo nets 45 cm mouth diameter with 330 micron mesh zooplankton net equipped with a calibrates flowmeter at the mouth to measured volume of water filtered. The tow were made from about 150 m depth (mean depth=142 m, S.D.=18.9 m) to the surface at a speed of approximately 2 to 3 knots.

The samples were fixed in 5% formalin seawater solution and bring back to laboratory for sorting and transfer to 50% isopropyl alcohol for permanent preservation.

Cephalopod paralarvae were identified to the lowest possible taxa with the aid of published figures and descriptions of Okutani and Mcgowan (1969); Sweeney *et al.* (1992); Wakabayashi (1993) and Jivaluk (2001).

Dorsal mantle length (mm ML) of all undamaged paralarvaes was measured to the nearest 0.1 mm with an ocular micrometer connected to a stereomicroscope.

The number of cephalopod paralarvae per tow was standardized to number of paralarvae per $1,000 \text{ m}^3$ (individuals/1,000 m³).

Small paralarvae of families Ommastrephidae and Onychoteuthidae could be distinctive identified only at family level. The identification was not possible either genus or species level because a growth series connecting paralarvae to known adults were not available.



Figure 1 Map of sampling stations of the cephalopod paralarvae in the Bay of Bengal.

Results

Captured Composition

A total of 278 paralarvae of 13 families and 19 genera was collected from 29 of 34 stations (Table 1). The most numerous families were the Ommastrephidae (41% of total

catch) followed by families Enoploteuthidae (14%) and Onychoteuthidae (6%). The majority of individuals were *Nototodarus hawaiiensis* (49 individuals) and *Abraliopsis* sp. (24 individuals).

Only a small number of 24 squids from 9 genera of other families (Ancistrocheiridae, Brachioteuthidae, Chiroteuthidae, Chtenopterygidae, Cranchiidae, Histioteuthidae, Octopoteuthidae, and Thysanoteuthidae) and 6 octopods (order Octopoda; suborder Incirrata) of 2 genera from family Bolitaenidae and Octopodidae were captured (Table 1).

Distribution and Abundance

Order Teuthida (Suborder Oegopsida)

Family Ommastrephidae was the most widely distributed, occurring at 26 stations (Fig. 2). *Nototodarus hawaiiensis* was the most numerous species collected, comprising 43% of the ommastrephid catch, occurring at 13 stations of the survey area A and B (Fig. 3). *Sthenoteuthis oualaniensis* was occurred in survey area A, B and C, collected at 9 stations (Fig. 4).

Family Enoploteuthidae was collected at 18 stations and also occurred in every survey areas (Fig. 5) but less abundance than family Ommastrephidae. *Abraliopsis* sp. was the highest number collected at 11 stations, composed 25% of enoploteuthid catch and more abundance in area B (Fig. 6). *Abralia* sp. and *Enoploteuthis* sp. collected at 4 and 3 stations, occurring in the survey area B and C, but *Enoploteuthis* sp. was less abundance than those of *Abralia* sp. (Figs. 7 and 8).

Family Onychoteuthidae was less abundance than family Ommastrephidae and Enoploteuthidae, occurring at 8 stations in survey areas A, B and C (Fig. 9). *Onychoteuthis* sp. was collected at 3 stations of survey area A and B and composed 25% of onychoteuthid catch (Fig. 10). Onychoteuthid species A were collected only in survey area C, composed 75% of onychoteuthid catch (Fig. 11).

Family Chtenopterygidae contained 9 individuals of *Chtenopteryx* sp. (3% of total catch) were collected at 5 stations of area B and C (Fig. 12).

Family Chiroteuthidae was collected at 5 stations from area A, B and C (2% of total catch). *Chiroteuthis* sp. occurred in three areas but *Asperoteuthis* sp. was found a single specimen in area B (Fig. 13).

Family Brachioteuthidae was collected 3 individuals of *Brachioteuthis* sp. at 2 stations of area B and C (Fig. 14).

Family Thysanoteuthidae was collected only 2 individuals of *Thysanoteuthis rhombus* at 2 stations of area A and C (Fig. 15).

Single specimen of 4 genera (*Ancistrocheirus lesueuri*, *Liocranchia* sp., *Histioteuthis* sp., *Octopoteuthis* sp.) from 4 families (Ancistrocheiridae, Cranchiidae, Histioteuthidae, Octopoteuthidae) were found in area B and C (Table 1).

Order Octopoda (Suborder Incirrata)

Octopus sp. (family Octopodidae) was collected at 2 stations (3 individuals) of area C and 1 station (1 individual) of area A. Only 2 individuals of *Japettella* sp. (family Bolitaenidae) was collected at the same station (station 23) in area A (Fig. 16).

Table 1 Number of species and individuals of cephalopod paralarvae collected by zooplankton net during the BIMSTEC survey and

| mantle] | engt | h rí | ang | je (r | nm | () O | f sa | dur | oles. | | | | | | | | | | | | | | | | | | | | | | | | | |
|---|------------|------|-----|-------|----|------|------|-------------|-------|----------------|--------|-----------|------|----|--------------|------|--------------|-------------|------|-------|------|----|----|----|----|----|------|--------|--------|----|----|------------------|-----------------------|-------------------------|
| | | | | | | Area | S | | | | | | | | | | | Are | аA | | | | | | | | | Are | аB | | | | MLra | ange |
| | - | 7 | е | 4 | 5 | 9 | 7 | 8 | 6 | 10 | 11 1: | 2 | 3 14 | 15 | 16 | 17 1 | 18 1 | 9 2 | 0 21 | 22 | 23 | 24 | 25 | 26 | 27 | 28 | 29 3 | 0 3 | 1 32 | 33 | 34 | Total | m) | E |
| Order Octopoda Family Bolitaenidae <i>Japettel</i> la sp.? | | | | | | | | | | | | | | | | | | | | | р | | | | | | | | | | | 2 | 2.3 - | - 8.5 |
| Family Octopodidae Octopus sp. | | | 2 | | | | | - | | | | | | | | | | | | | | | | | ~ | | | | | | | 4 | - 1.7 - | - 3.9 |
| Order Teuthidae Family Ancistrocheiridae Ancistrocheiru s lesueuri | | | | | | | | | - | | | | | | | | | | | | | | | | | | | | | | | - | | |
| Family Brachioteuthidae Brachioteuthis sp. | 2 | | | | | | | | | | | | | | | | | | | | | | | | | | | - | _ | | | ° | 3.5 - | - 4.2 |
| Family Chiroteuthidae Asperoteuthis sp. Chiroteuthis sp. | ~ | | | | | | | | | | | | ~ | | | | ~ | | | | | | | | | | | - | 7 | | | 5 - - | 3.9 - | - 6.9 |
| Family Chtenopterygidae <i>Chtenopteryx</i> sp. | | | ~ | | | | | | | | | | | | | | | | | | | | | | | | | 2 | 4 | - | | 6 | - 6:0 | - 4.5 |
| Family Cranchiidae <i>Liocranchi</i> a sp. | - | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | - | | |
| Family Enoploteuthidae Abralia sp. Abraliopsis sp. Frondintarthis sp. | <i>с</i> с | ~ | | | | Ν | | | N | , γ | | | | | ~ | | (1) | 5 | - | | | ~ | | | | | ~ | | - + | ~ | Ν | 11 24 | 1.8 - 1.4 - 2 | - 7.2 - 5.6 - 5.1 |
| Family Histioteuthidae Histioteuthis sp. | | | | | | | | | ~ | | | | | | | | | | | | | | | | | | | | | | | , , | | i |
| Family Octopoteuthidae Octopoteuthis sp. | | | | - | | | | | | | | | | | | | | | | | | | | | | | | | | | | - | | |
| Family Ommastrephidae Sthenoteuthis oualaniensis Nototodarus hawaiiensis? Ommastrephid species | ~ ~ | 0 0 | - | ы | | e | | ~ | 2 | | с С | | | 0 | 7 10 | 4 0 | - | n a | 0 0 | 7 7 3 | œ | з | ~ | р | | | ~ | - ~ | ю _ | | 10 | 20 49 46 | 2.0 0.8 - 1.4 - | - 5.1 - 2.6 - 2.1 |
| Family Onychoteuthidae Onychoteuthis sp. Onychoteuthid species A | с | 9 | | - | | | | - | - | | | | | | | | | | - | | - | | | | | | | | | | 2 | 4 12 | 2.4 - 1.7 - | - 7.4 - 2.9 |
| Family Thysanoteuthidae Thysanoteuthis rhombus? | | | - | | | | | | | | | | | | | | | | | | - | | | | | | | | | | | 2 | | |
| Unidentified | 12 | 9 | - | 12 | | - | | 9 | 5 | 5 | 5 2 | <u>c'</u> | - | | | | | | - | | | | | - | | | 4 | 3 7 | - | | | 79 | 0.5 - | 10.8 |
| Total cephalopods | 27 | 17 | 9 | 17 | 0 | 9 | 0 | 16 | 18 | 6 | 95 | 5 | 0 2 | 2 | 4 | 7 | - | 9 | 14 | 1 17 | . 12 | 4 | 7 | 3 | - | 0 | 181 | 0 | 6 10 | 4 | 8 | | | |



Figure 6 Distribution and abundance (ind./1,000m³) of *Abraliopsis* sp.



Figure 3 Distribution and abundance (ind./1,000m³) of *Nototodarus hawaiiensis*.



Figure 5 Distribution and abundance(ind./1,000m³) of family Enoploteuthidae.



Figure 7 Distribution and abundance (ind./1,000m³) of *Abralia* sp.



Figure 12 Distribution and abundance (ind./1,000m³) of *Chtenopteryx* sp.



Figure 9 Distribution and abundance (ind./1,000m³) of family Onychoteuthidae.



Figure 11 Distribution and abundance (ind./1,000m³) of Onychoteuthid species A.



Figure 13 Distribution and abundance (ind./1,000m³) of *Asperoteuthis* sp. and *Ancistrocheirus lesueuri*.



Figure 14 Distribution and abundance (ind./1,000m³) of *Brachioteuthis* sp.



Figure 16 Distribution and abundance (ind./1,000m³) of *Octopus* sp. and *Japettella* sp.



Figure 15 Distribution and abundance (ind./1,000m³) of *Thysanoteuthis rhombus*.

Discussion

The results of the present study showed that the paralarvae catch by zooplankton net during the survey are all oceanic species of various families of suborder Oegopsida (order Teuthida) except 2 families from suborder Incirrata (order Octopoda). The benthic adult octopods (Octopus sp.) were found occurred more than 200 km offshore of the Myanmar and Bangladesh waters. The results of the distribution found the assemblages of the pelagic adults in the continental slope and oceanic waters (Abralia sp., Abraliopsis sp., Enoploteuthis sp.), epi-mesopelagic adults (Sthenoteuthis oualaniensis, Nototodarus hawaiiensis, Onychoteuthis sp. Thysanoteuthis rhombus), mesopelagic adults (Asperoteuthis sp., Chiroteuthis sp., Histioteuthis sp.), and meso-bathypelagic adults (Chtenoteryx sp., Liocranchia sp., Octopoteuthis sp., Brachioteuthis sp.). A diverse community and the most numerous numbers of captured occurred in the survey area off Myanmar waters. Especially, the abundance of the small size of Ommastrephid species (<2 mm) suggesting the nearshore spawning in this area. Possibly the good feeding grounds influenced by surface water runoffs transporting nutrientrich freshwater into the coastal areas (Sundström et al., 1987; Janecarn and Chullasorn, 1997; Limpsaichol et al., 1998) and/or upwelling conditions create by warm surface waters mixing the nutrient rich bottom (Dwivedi and Choubey, 1998). As the inshore-offshore spawning migrations is common in many cephalopod species (Nesis, 1993a; Mangold, 1987). Some *Enoploteuthis* species have also been reported to spawn only over the slopes or in nearshore oceanic regions (Nesis, 1993a and 1996). It is possible that spawning of ommastrephid squids may occur throughout the Bengal Bay. The composition from the results also show that the abundant paralarval squid is the commercial and/or minor-commercial target species to fisheries (Carpenter and Niem, 1998).

The results from the present study is useful for determining when and where adults spawn, particularly for species whose adults are difficult to catch (Bower *et al.*, 1999). Further sampling survey in different monsoon season will help to provide a better understanding on life history of cephalopod species in these areas.

Acknowledgements

The authors wish to acknowledge the cooperation between the BIMSTEC member countries, Department of Fisheries of Thailand and SEAFDEC/TD. We would also like to thanks to the officers and crews of M.V. SEAFDEC for their helps during the surveys.

References

- Arkhipkin, A. I., Y. N. Zheronkin, Y. A. Loktionov and A. S. Schetinnikov. 1988. Fauna and distribution of pelagic cephalopods larvae in the Gulf of Guinea. *Zool. Zh.* 67(10):1459-1467. (in Russian, English abstract)
- Bower, J. B., M. P. Seki, R. E. Young, K. A. Bigelow, J. Hirota and P. Flament. 1999. Cephalopod paralarvae assemblages in Hawaiian waters. *Mar.Eco.Prog.Ser.* 185:203-212.
- Carpenter, K. E. and V. H. Niem. (eds.). 1998. FAO species identification guide for fishery purposes. The living marine resources of the Western Central Pacific, vol. 2, Cephalopods, crustacean, holothurians and sharks. FAO Publication, Rome. p. 764-780.
- Dwivedi, S. N. and A. K. Choubey. 1998. Indian Ocean large marine ecosystems: need for national and regional framework for conservation and sustainable development.In: K Sherman, K., E. Okemwa and M. Ntiba. (eds.). Large Marine Ecosystems of the Indian Ocean: Assessment, Sustainability and Management. Blackwell Science.
- Janekarn, V. and S. Chullasorn. 1997. Environmental impacts on coastal fisheries along the west coast of Thailand. In: Asia-Pacific Fishery Commission (APFIC): Environmental aspects of responsible fisheries. Proceedings of the APFIC Symposium Seoul, the Republic of Korea, 15-18 October 1996. FAO Bangkok. RAP Publication 32/1997. p. 222-233.
- Jivaluk, J. 2001. Species, abundance and distribution of cephalopod paralarvae in the Gulf of Thailand in 1995 and 1996. *Fish. Mus. Nat. Hist. Tech. Pap. No. 1/2001.* 31 pp.
- Kubodera, T. 1991. Distribution and abundance of the early life stages of octopus, *Octopus dofleini* Wulker 1910 in the North Pacific. *Bull. Mar. Sci.* 49(1-2):235-243.
- Kubodera, T. and K. Jefferts. 1984a. Distribution and abundance of the early life stages of squid, primarily Gonatidae (Cephalopoda: Oegopsida) in the northern North Pacific (Part 1). Bull. Natl. Sci.Mus. Ser. A. (Zool). 10(3):91-106.
- Kubodera, T. and K. Jefferts. 1984b. Distribution and abundance of the early life stages of squid, primarily Gonatidae (Cephalopoda: Oegopsida) in the northern North Pacific (Part 2). Bull. Natl. Sci.Mus. Ser. A. (Zool). 10(4):165-193.
- Kubodera, T. and T. Okutani. 1981. The systematics and identification of larval cephalopods from the northern North Pacific. *Res. Ins. N. Pac. Fish. Hokkaido Univ. Spec.* p. 131-159.
- Limpsaichol, P., S. Khokiattiwong, N. Bussarawit and P. Sojisuporn. 1998. Environmental factors influencing the health and productivity of the Phang-nga Bay. In: Communitybased fisheries management in the Phang-nga Bay, Thailand. Proceedings of the National Workshop on Community-based Management organized by the Department of Fisheries of Thailand, FAO and the Bay of Bengal Programme, Phuket, Thailand, 14-16 February 1996. Phuket, Thailand. p. 85-120.
- Mangold, K. 1987. Reproduction. **In:** Boyle PR. (ed). Cephalopod Life Cycles. Academic Press Inc, London. p. 157-200.
- Nesis, K. N. 1974. Cephalopod larvae in the western Arabian Sea. *Oceanology* 14(3): 441-445.
- Nesis, K. N. 1993. Cephalopods of seamounts and submarinendges. In: Okutani, T., R. K. O'Dor and T. Kubodera, T. (eds.). Recent Advances in Cephalopod Fisheries Biology. Tokai University Press, Tokyo. p. 365-373.
- Nesis, K. N. 1996. Mating, spawning and death in Oceanic cephalopods: a review. *Ruthenica* 6(1):23-64.
- Okutani, T. 1966. Studies on early life history of decapodan mollusca II Planktonic larvae of decapodan cephalopods from the northern North Pacific in summer seasons during 1952-1959. *Bull. Tokai. Reg. Fish .Res. Lab.* 45:61-79.

- Okutani, T. 1968. Studies on early life history of decapodan mollusca-III. Systematics and distribution of larvae of decapod cephalopods collected from the sea surface on the Pacific coast of Japan 1960-1965. *Bull. Tokai. Reg. Fish. Res. Lab.* p. 559-57.
- Okutani, T. 1969. Studies on early life history of decapodan Moliusca-IV. *Bull. Tokai. Reg. Fish. Res. Lab.* 58:83-96.
- Okutani, T. and J. A. McGowan. 1969. Systematics, distribution, and abundance of the epiplanktonic squid (Cephalopoda: Decapoda) larvae of the California Current Aprd 1.954-March 1957. *Bull. Scripps. Inst. Oceanogr. Univ. Calif.* 14:1-90.
- Piatkowski, U. and W. Welsch. 1991. On the distribution of pelagic cephalopods in the Arabian Sea. *Bull. Mar. Sci.* 49(1-2):186-198.
- Piatkowski, U., W. Welsch and A. Ropke. 1993. Distribution patterns of the early life stages of pelagic cephalopods in three geographically different regions of the Arabian Sea.
 In: Okutani, T., R. K. O'Dor and T. Kubodera. (eds.). Recent Advances in Cephalopod Fisheries Biology. Tokai University Press, Tokyo. p. 417-431.
- Saito. H. and T. Kubodera. 1993. Distribution of *Ommastrephid rhynchoteuthion* paralarvae (Mollusca: Cephalopoda) in the Kuroshio region. In: Okutani, T., R. K. O'Dor and T. Kubodera. (eds.). Recent Advances in Cephalopod Fisheries Biology. Tokai University Press, Tokyo. p. 457-466.
- Sundström, B., V. Janekarn, J. Hylleberg and P. Boonruang. 1987. Annual pelagic primary production with notes on physical and chemical variables at Phuket, the Andaman Sea, Thailand. *Res. Bull. Phuket Mar. Biol. Cent.* 46:12 pp.
- Sweeney, M. J., C. F. E Roper, K. M. Mangold, M. R. Clarke, and S. V. Boletzky. (eds.). 1992. Larval and Juvenile Cephalopods: A Manual for their Identification. Smithson. Contrib. Zool. 513:282 pp.
- Vecchione, M. 1987. Juvenile ecology. **In:** Boyle, P. R. (ed.). Cephalopod Life Cycles, Vol. 2. Academic Press Inc, London. p. 61-84.
- Wakabayashi, T. 1993. Species, distribution and abundance of cephalopod paralarvae in the western Australian waters. Master Thesis. Tokyo University of Fisheries, Tokyo. 120 pp.
- Yamamoto, K. and T. Okutani. 1975. Studies on early life history of decapodan mollusca-V. Systematics and distribution of epipelagic larvae of decapod cephalopods in the southwestern waters of Japan during the summer in 1970. *Bull. Tokai Reg Fish Res. Lab.* 83:45-96.
- Young, R. E. and R. F. Harman, R. F. 1988. Larvae, paralarvae and subadult in cephalopod terminology. *Malacologia* 29:201-207.